

ANTHOCYANIN STABILITY OF ROBUSTA COFFEE CHERRIES DURING STORAGE

Sukatiningasih¹, Windrati, W.S, and Yudistira D

Abstract

The overripe Robusta cherries had potential as a source of anthocyanin. The aim of this research was to investigate anthocyanin stability which extracted from cherries. Anthocyanin was extracted using ethanol and aquadest with ratio 1:1. Extraction was repeated 3 times until the residue was colorless. After that this extract was kept at different temperatures 5, 25 and 45 °C for 4 weeks. The stability was evaluated by content and lost anthocyanin, Hue color, pH and antioxidant activity. The result showed that during storage anthocyanin stability of all samples decreased. Sample which kept at 5 °C had the best stability with anthocyanin content 3.26 %, lost anthocyanin 26.55 %, Hue color 146.12, pH 5.01 and antioxidant activity 16.26 % after 4 weeks. The least stability was obtained at sample which kept at 45 °C with anthocyanin content 1.30 %, lost anthocyanin 29.02 %, Hue color 124.92, pH 5.03 and antioxidant activity 6.60 %.

Keywords : *Robusta cherries, anthocyanin stability, antioxidant activity, temperature*

Introduction

Anthocyanins mainly contribute to the bright red color of fruits, vegetables, and grains [3]. Epidemiological data associate anthocyanins with prevention of various diseases such as visual and vascular diseases such as vascular diseases, obesity and some cancers [18; 8; 17; 1; 13; 6].

Recently, as consumers are increasingly concerned about the safety of synthetic colorants used in food, the trend of using natural colorants in food products has also increased intensively. The potential health benefits of anthocyanins enhance consumer interest in using it in food to replace artificial red or purple colorant [12]. Anthocyanin colorant could be used in many solid foods such as extruded snacks and baked cakes as well as in drinks and beverages [11].

The degradation of the anthocyanins in fruits and vegetables during processing and storage has been reported in some studies [9; 14; 11]. Those studies were carried out at a temperature below 100°C and a high moisture condition. The degradation of the anthocyanins was significantly accelerated by various glycosidase activities and severe pH in the sample, beside heating temperature. Thus, the liability of the anthocyanins observed in those studies could be different from that under dry heating condition.

¹ Faculty of Agricultural Product Technology, Jember University

The overripe Robusta coffee cherries containing anthocyanin 8.34 % , pH 5 showed the highest stability during storage [16].

At pre-research, have been tried to get Robusta coffee extract by using aquadest: ethanol (1:1) as a solvent. Robusta coffee cherries extract solution preparation. In this study, robusta coffee cherries extract was used to evaluate the stabilities of anthocyanin at different temperature during storage. Also antioxidant activities of anthocyanin was evaluated using DPPH (2,2'-diphenyl-1-picrylhydrazyl) free radical scavenging method as well.

Material and Methodes

Raw material which used in the making of extract of robusta coffee is the fruit of overripe Robusta coffee with red colour from Durjo estate, Jember, East Java Indonesia. We also use glicerol and Citric acid

Chemicals and Equipment

Aquades, ethanol 97%, Reagent DPPH, acetic acid, buffer pH 1 and pH 4.5, NaOH. Equipment needed in this research were rotavapor, blender, cloth filter, glassware, Eppendorf micropipete, pH metre, oven, centrifuge, cooler.

Statistical Analysis

Design research were consist of 2 factors. Three times replication to every treatment. D= temperature of storage D1= 5°C, D2 = 25 °C and D3= 45°C and length of storage M: M1= 1 week, M2= 2 weeks, M3=3 weeks and M4= 4 weeks. The data was analyzed by Descriptif test and showed the histogram.

Overripe Robusta coffee cherries extract:

After washing the fruit, separated the pulp and seed. 15 grams the pulp blend 75 ml solvent (contained of aquadest : ethanol 97% 1:1 and 15% citric acid v/v), maceration during 15 minute and filtered the solution. The residue replicated extracted 2 times until colorless. Then centrifugated on 4000 rpm during 20 minutes. After that the filtrate to concentrated by rotary evaporator on 40°C until the volume of extract 75 ml. The extract made pH 5 and then added with 10% glicerol. The sample packing on the bottle and then kept in 5°C, 25°C, 45°C and storage for 4 weeks.

Total Anthocyanin with pH differential method (Giusti and Wrolstad, 2001)

Total Anthocyanin concentration : Dissolved 1 ml sample with 4 ml KCl (pH 1)) for 20 min and Na-acetate (pH 4,5) for 20 min. Measured every solution in 520 nm and 700 nm with buffer pH 1 and buffer pH 4,5 as standard.

$$A = (A_{520} - A_{700})_{\text{pH 1}} - (A_{520} - A_{700})_{\text{pH 4,5}}$$

Total anthocyanin formula :

$$\square \text{otal anthocyanin} = \frac{A}{\epsilon \times L} \times \text{MW} \times \text{DF} \times \frac{V}{\text{Wt}} \times 100\%$$

Explanation

ϵ = Absorptivity molar Sianidin-3-glukoside = 26900/mol

L = Kuvet width = 1 cm

M MW=Molecule weight of Sianidin-3-glukoside = 449.2 g/mol

DF = Dilluted factor

V = Volume (L)

Wt = Sample weight (g)

Detemination Lost Anthocyanin

$$\text{Lost Anthocyanin (\%)} = \frac{\text{mg/l anthocyanin (a-b) } \times 100\%}{\text{Mg/l anthocyanin a}}$$

Explanation :

a = concentration anthocyanin before storage

b = concentration anthocyanin after storage

Determination of color (Color reader, Fardiaz et al, 1992)

The initial color used ceramic as standard had L. A.b value sequently 94.35; -5,75 and 6.51

$$H = \tan^{-1} \frac{b^*}{a^*}$$

Explanation :

a * = range value -80 – (+ 100), showed green until red

b * = range value – 50 – (+70), showed blue until yellow

H = angel of color (0°= neutral color.90° = yellow. 180° = green.270° = blue)

Determination pH (pH metre)

Investigated pH using pH meter, the electrode immersed on buffer pH 7, after that immersed on the sample until stable then noticed.

Determination of DPPH free radical scavenging capability (Gadow, 1997)

The DPPH free radical capability in the robusta coffee extract was determined using the spectrophotometer method. The robusta coffee extract 10 µl in the test tube was redissolved in 1 ml DPPH for 20 min, added with ethanol 97% until 5 ml, vortex measured the absorbance on 517 nm. The concentration of DPPH solution was 0.0394 g in 250 ml ethanol 97% (the concentration 400 µ mol/l). Kurva standard DPPH $Y = 1.791x + 0.014$
Aktivitas antioxidant = $\frac{A(\text{blanko} - \text{sample})}{A(\text{blanko})} \times 100\%$

A blanko

Explanation

Blanko made from DPPH solution without sample

Y = absorbance

X = konsentrasi (µ mol DPPH)

Result and Discussion

Anthocyanin content

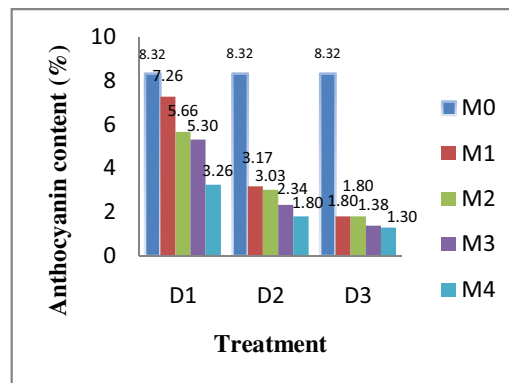


Figure 1. Anthocyanin Content of Robusta Coffee Cherries Extract

Histogram of anthocyanin Robusta Coffee Cherries extract is shown in Figure 1. The total anthocyanin content initially is 8.32%. The highest content anthocyanin in the extract was D1 (7.26% this extract kept on 5 °C the first week, 5.66% on the second weeks, 5.30% on the three and 3.26% the fourth weeks). In this study shown the anthocyanin

more stabil kept on 5 0C .(6), investigated the characteristic of anthocyanin which kept on various temperature, the result shown the anthocyanin degradation more so much with the temperature hight . The high temperature accelerated the degradation of anthocyanin.(8;13).The range of total anthocyanin content was from 1.298 % to 7.26 %.

Lost anthocyanin

Lost anthocyanin Robusta Coffee Cherries extract shown in Figure 2. The highest lost anthocyanin shown on D3 it was 29.02 %.This shown the temperature storage high so much, the degradation anthocyanin also so much. The range of degradation is was 27.05 % to 29.02 %

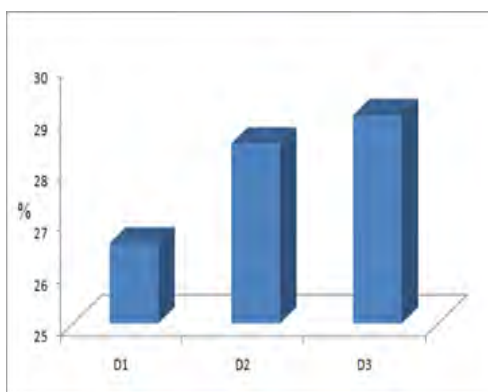


Figure 2.Lost Anthocyanin of Robusta Coffee Cherries Extract

Color of Robusta Coffee Cherries.

The color of robusta coffee cherries shown in Figure 3 and Figure 4. Figure 3 shown robusta coffee cherries extract kept on 5⁰C (D1) had bright red then extract which kept on 25⁰C had purple red and extract kept on 45⁰C had dark red. The finished observation extract kept on 45⁰C the color so much dark beside extract kept on 27⁰C and 5⁰C so much purple.

The observation on H value shown on Figure 4. Range of H value were 129.962 to 155.028. There were Yellow Green (YG) , beside Mo had value 144.382 (data not presented). There were shown on the same range. Maybe the sample contained the other substance as ascorbic acid, polifenol, carotenoid and the others so that color combined shown Yellow Green.

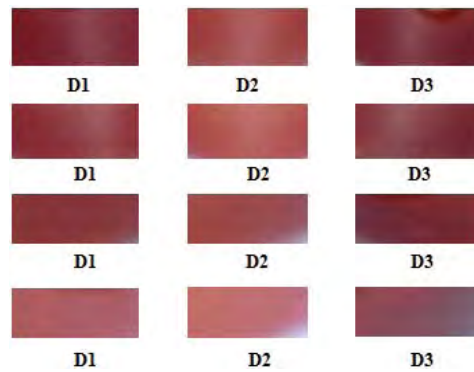


Figure 3. Color of Robusta Coffee Cherries Extract

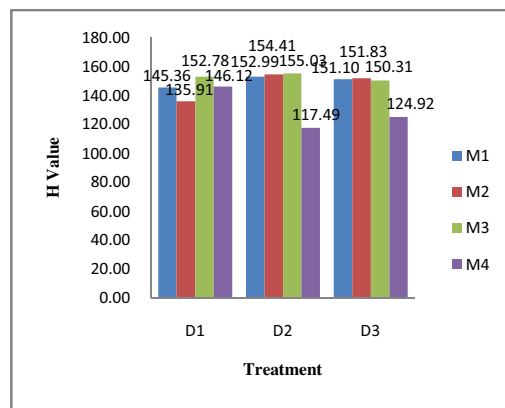


Figure 4. Color (Hue) of Robusta Coffee Cherries Extract

pH

Figure 5 shown pH robusta coffee cherries extract on various treatment. Stored treatment on 5⁰C; 25⁰C; 45⁰C shown the pH increased. Range of pH 4.73 to 5.04. The Initial pH 4.81 (data not presented), there were shown during storage pH extract trend increased on all the treatment. There were shown degradation anthocyanins also happened on all sample. Degradation of anthocyanin result increased pH

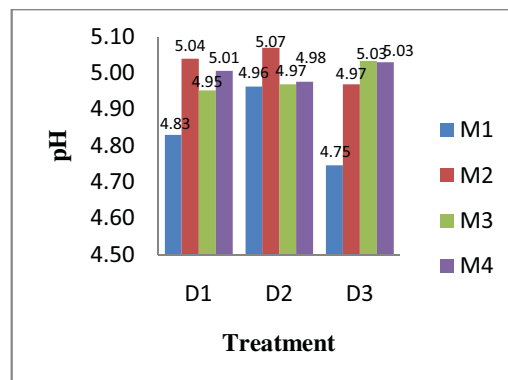


Figure 5. pH of Robusta Coffee Cherries Extract

Antioxidant Activity

The changes of DPPH free radical scavenging capability of the robusta coffee cherries extract shown on Figure 6. The highest antioxidant activity shown on D1 treatment (extract kept on 5⁰C).The range of antioxidant activity 6.60 % to 24.39 %. M0 (before storage) had antioxidant activity 78.16 %. There were shown antioxidant activity had trend decreased during storage all the treatment. But the decreased highest was the sample wich kept on 45 ⁰C. There were shown high temperature accelerated the degradation antioxidant substance include anthocyanin. It this as regard [10] reported radical scavenging activity of acylated anthocyanin in the red radish extract. Also, some degradation product of anthocyanins were reported to have antioxidant capability [15]

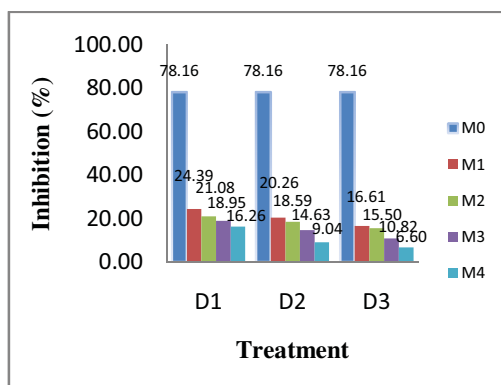


Figure 6. Antioxidant Activity (Inhibition %) of Robusta Coffee Cherries Extract

Conclusions

From this study, stabilities of anthocyanin during storage were revealed. The anthocyanin trend decreased during storage, there were not stable on storage in low temperature, the decreased it small but on high temperature was as high as the temperature. The optimal temperature storage it 5⁰C, anthocyanin content was 3,34 %, beside on 25⁰C and 45⁰C there were 1.80 % and 1.30 % after 4 weeks.

References

- [1].Canter PH, Ernst E. Anthocyanides of *Vaccinium myrtillus* (blueberry) for night vision – a systemtic review of placebo –controlled trials. *Surv Ophtalmol* 4938-50, 2004
- [2].Fardiaz, D., N.Andarwulan, H.Wijaya and N.L. Puspitasari. *Technology of Analysis Chemistry and Functional Property Component of Food. Laboratorium Guide PAU Food and Nutrition, IPB Bogor, 1992.*

- [3].Francis F J.Food Colorings.In Mac Dougall DB, Editor, Color in Food, Boca Raton, Fla CRC Press.p 297-330, 2002
- [4].Gadow,A,Joubert,E and Hansman,CF. Comparison of Tea Antioxidant Activity of Aspalatin with that of Plant Phenols of Rooibos Tea (*Aspalathus linearis*).J Agric Food Chem. 45:832-638.1997
- [5].Giusti, M.M. and R.E. Wrolstrad. Characterization and measurement of Anthocyanin by UV Visible Spectroscopy. Current Protocol in Food Analytical Chemistry. 2001http://www.nshtvn.org/e_book/molbio/current_protocols/CPFA/faf_0102.pdf.
- [6].Jang YP, Sparrow JR, Zhou J, Nakanishi K. Anthocyanins protect against A2E photooxidation and membrane permeabilization in retinal pigment epithelial cells. Photochem Photobiol 81 529-36 .2005
- [7].Kampuse S,K.Kampuss,L.Pizika. Stability of Anthocyanins and Ascorbic Acid in Raspberry cultivar During Frozen Storage.in Musa 2006.Natural Coloring from Duwet (*Syzium cuminii*) Study The Production and Stability During Storage. Faculty of Agricultural Technology UNEJ.2005
- [8].Katsube N, Iwashita K.,Tsushida T.,Yamaki K.,Kobori M.Induction on apoptosis in cancer cells by blueberry (*Vaccinium myrtyllus*) and the anthocyanins.J.Agric.Food Chem.51 68-75, 2003
- [9].Kirca A., Ozkan M.,Chemeroğlu B. Stability of black carrot anthocyanins in various fruit juices and nectars. Food Chem. 97 598-605, 2006
- [10].Matsufuji H, kido H, Misawa H, Yaguchi J, Otsuki T, Makoto C, Misuharu T, Yamagata K.Stability of liht, heat and hydrogen peroxide at different pH values and DPPH radical scavenging activity of acylated anthocyanins from red-radish extract.J.Agric Food Chem 55 3692-701.2007
- [11].Mishra DK., Dolan KD.,Yang L. Confidence interval for modeling antocyanin retention in grape pomace during nonisothermal heating.J.Food Sci 73 E9-14.2008
- [12].Nielsen SR and Holst S. Developments in natural coloring, In Mac Dougall DB.editor.Color in Food. Boca Raton, Fla CRC press, p.331-51. 2002
- [13].Prior RL., Joseph J. Berries and fruits in cancer chemoprevention. In Bagchi D, Preuss HG, edition. Phytopharmaceuticals in cancer chemoprevention Boca Raton, Fla CRC Press, p.465-79 . 2004
- [14].Sadilova E., Stintzing EC., Carle R. Thermal Degradation of acylated and nonacylated anthocyanins.j.Food Sci 71 C504-12.2006
- [15].Seeram NP, Bourquin LD, Nair MG.. Degradation products of cyaniding glycosides from tart cherries and their bioactivities.J Agric Food Chem 49 4924-9. 2001

[16].Sumartiningdha,R.A. Study Rapening and solvent on extraction of antioxidant substances Robusta Coffee Cherries. FTP UNEJ, Jember 2011

[17].Tsuda T, Horio F,Uchida K,Ozawa T. Dietary Cyanidine 3-O-b-D glucoside-rich purple corn color prevents obesity and meliorates hyperglycemia in mice.J Nutr.133 2125-30.2003.

[18].Youdim KA,Mc Donald J,Kalt W,Joseph JA.Potential role of dietary flavonoids in reducing microvascular endothelium vulnerability to oxidative and inflammatory insults,J.Nutr.Biochem 13282-8.2002